

GYRATE ATROPHY-LIKE PHENOTYPE WITH NORMAL PLASMA ORNITHINE

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Purpose: To describe the clinical characteristics of a chorioretinal disease with a gyrate atrophy-like phenotype and normal plasma ornithine.

Methods: One family with three men who had progressive chorioretinal disease and three additional patients with simplex cases were examined clinically and with standard electroretinography, electrooculography, and dark adaptometry.

Results: In the family, a 70-year-old man and his two sons (39 and 41 years of age) were affected. On ophthalmoscopy, sharply demarcated peripheral patches of retinal pigment epithelium and choroidal atrophy were seen to progress to the posterior pole in the father's eye. In three unrelated men (62, 70, and 80 years of age), chorioretinal atrophy was present in the mid- and far periphery. Visual acuity was normal in the two youngest of all six patients; however, electroretinogram and electrooculogram waves were reduced. Advanced visual field defects and visual acuity loss occurred in the four older patients. Electroretinogram and electrooculogram were reduced, and the dark adaptation thresholds were elevated. In all patients, serum ornithine levels were normal. Ornithine-delta-aminotransferase activity in cultured skin fibroblasts and the apparent Michaelis constant (Km) for ornithine and α -ketoglutarate were within the normal range in all patients.

Conclusions: A gyrate atrophy-like phenotype can result from causes other than deficient ornithine-delta-aminotransferase. Its occurrence in three male members in two generations in one family suggests an autosomal dominant inheritance in at least some such patients.

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Gyrate atrophy is an autosomal recessive chorioretinal dystrophy.¹⁻¹³ Most patients with this disorder have high myopia, and posterior sub-

capsular cataracts are common. The chorioretinal atrophy starts in the midperiphery and proceeds to the periphery and toward the posterior pole. Later, the dystrophic areas become confluent. There is a corresponding progressive constriction of visual fields. For these patients, severe visual acuity loss usually occurs in the fifth or sixth decade of life. Patients affected with gyrate atrophy show various mutations in the gene encoding the enzyme ornithine-delta-aminotransferase (OAT).¹⁴⁻²² These mutations lead to markedly elevated serum levels of ornithine. The diagnosis of gyrate atrophy can be established by the examination of serum ornithine levels or OAT enzyme activity in cultured skin fibroblasts of affected patients and carriers.^{4,23-31} Since the detection of hyperornithinemia in patients

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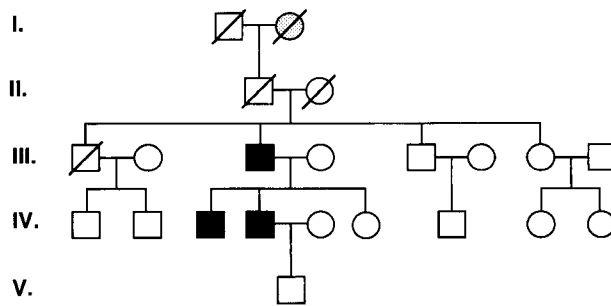


Fig. 1. Pedigree of family M. Black symbols indicate affected patients; grey symbols indicate patients who were probably affected.

with gyrate atrophy in 1973,²³ the term “gyrate atrophy” has been reserved for choroidal dystrophies that show hyperornithinemia.

Gyrate atrophy can be differentiated from chorioideremia and diffuse choriocapillaris atrophy based on clinical and biochemical findings and the inheritance pattern.^{32–36} Isolated, sporadic cases with ophthalmoscopic findings resembling gyrate atrophy and with normal serum ornithine levels have been described.^{37–43} Before the association between gyrate atrophy and hyperornithinemia was detected, individuals from two families who had chorioretinal dystrophy and autosomal dominant inheritance were diagnosed as having gyrate atrophy.^{33,44}

We observed one family with apparent autosomal dominant inheritance and three unrelated patients with a chorioretinal dystrophy resembling gyrate atrophy. All patients had normal serum ornithine levels. The purpose of this study is to describe the clinical and electrophysiologic findings in patients with this infrequently reported chorioretinal disease.

We studied six affected patients from three different eye departments. Electrophysiologic recordings were obtained according to the standard protocol for clinical electroretinography and clinical electrooculography.^{45,46} Details of the recording techniques have been published previously for each laboratory.^{47–50}

Case Reports

Family M

Four patients from Family M, including three affected men, were examined by one of the authors of this study (U.K.). The pedigree is shown in Figure 1. One patient (IV/4) was examined thoroughly in our department. His father (III/3) and brother (IV/3) were unavailable for examination in our department. Therefore, fundus examination and the drawing of blood samples were done at their home. The 6-year-old son (V/1) of patient IV/4 had normal visual acuity and fundoscopic appearance.

The paternal grandmother (I/2) of patient III/3 became blind at

approximately 50 years of age, but further details were not available. Both parents (II/1 and II/2) of patient III/3 died before the age of 45 years. The mother (III/4) of patients IV/3 and IV/4 had no difficulties with her vision; however, an examination was not possible. There was no positive family history for eye disease in the family of the mother (III/4). None of the other family members shown in the pedigree had known ocular problems; however, detailed ophthalmologic examination was not performed. The three affected men representing two generations and the woman who was probably affected suggest the presence of an autosomal dominant inheritance in Family M. In the second generation, no affected patient is known; however, the male family member II/1 may have died before clinical signs were present.

Patient IV/4. The male proband of the family had been diagnosed with retinitis pigmentosa when he was 18 years old. At that time, no subjective signs of ocular problems were present. The examination was performed because of the known disease in his father. His visual acuity was 20/20 in both eyes. Ophthalmoscopy showed a normal posterior pole and optic disc. Pigment epithelial changes were detected nasal to the disc and in the periphery. The vessels were described as normal. Goldmann visual fields were normal in both eyes (Figure 2). Electroretinography (ERG) and electrooculography (EOG) were recorded with unspecified techniques. The ERG results were described as slightly abnormal, and the EOG results revealed a severely reduced light rise.

This patient was examined by the authors (U.K. and M.H.F.) when he was 39 years old. At that time he had noted visual field defects. His general medical history was unremarkable, and he did not receive any medication. His visual acuity was unchanged. The refractive error was $+2.25-2.25 \times 11^\circ$ in the right eye and $+2.25-1.75 \times 172^\circ$ in the left. The anterior segments and the ocular pressures were normal. On ophthalmoscopy, patches of severe retinal pigment epithelial and choroidal atrophy in the mid- and far periphery were seen (Figure 3). These patches had become confluent in several areas. The borders between atrophic areas of the choroid and those that appeared to be normal were sharply delineated. For both eyes, there was a circular area of atrophy surrounding the optic disc, which temporally extended to approximately half the distance between the optic disc and fovea. The atrophic area around the disc was connected to the peripheral atrophy via a bridge of atrophic choroid and retinal pigment epithelium in the lower nasal quadrant. There was no hyperpigmentation or pigment clumping. Fluorescein and indocyanine angiography showed severe loss of choroidal vasculature with no leakage (Figure 3).

On visual field testing, concentric narrowing was detected on both eyes, and absolute paracentral scotomas (target V/4) were seen in connection with the blind spot (Figure 2). Results of color vision testing showed minor errors without typical axis in the desaturated Panel D-15 test. Results of tests using a Nagel anomaloscope were normal. Results of dark adaptation testing using a Goldmann-Weekers adaptometer with a target size of 11 degrees in the diameter in the upper field 10 degrees from the fovea showed normal cone and rod thresholds and a normal cone-rod break. Standard ERG recording showed a reduction of rod-mediated responses to approximately 15% and a reduction of cone-mediated responses to approximately 25% of the normal median; both were for responses mediated by photoreceptors (a-waves) and responses mediated by middle and inner retinal neurons (b-waves; Figure 4). Implicit times (the time from beginning of the light flash to the maximum of either the a- or b-wave in the ERG) were normal under dark and light adapted conditions. Standard EOG recording showed a reduced light rise

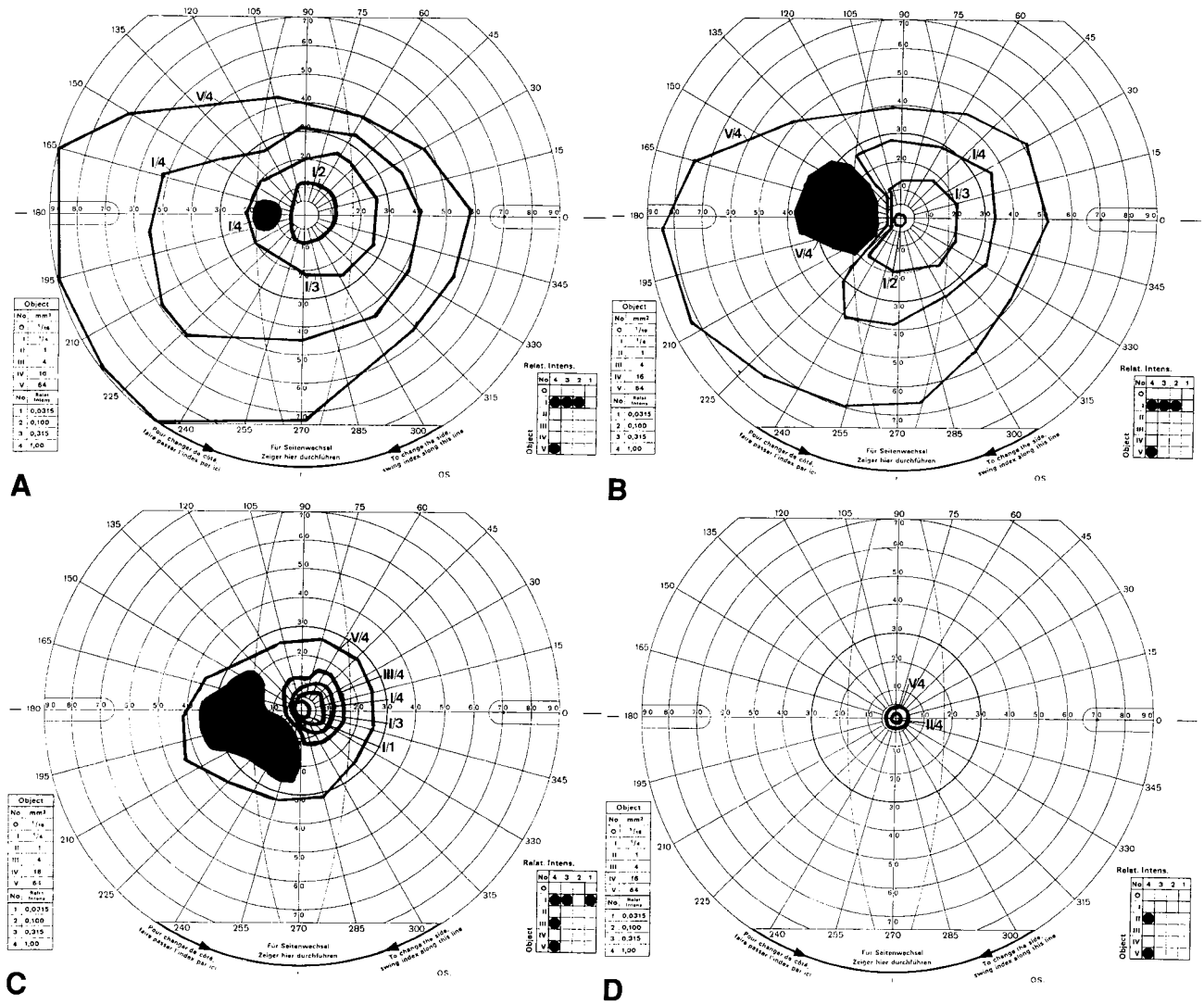


Fig. 2. Visual field defects in family M, patients III/3 and IV/4. **A**, Visual field of the left eye in patient IV/4 at 23 years of age. **B**, Visual field of the left eye in patient IV/4 at 39 years of age. **C**, Visual field of the left eye in patient III/3 at 51 years of age. **D**, Visual field of the left eye in patient III/3 at 62 years of age.

in both eyes (right eye, 1.28; left eye, 1.37; normal, > 1.60). The serum ornithine level was normal (122 $\mu\text{mol/L}$), as were the serum levels for other amino acids.

Patient IV/3. This 41-year-old healthy man had not noted his visual problems until his most recent examination. His visual acuity and visual fields had been tested once every year because he is a bus driver. He had been told that his visual acuity and visual fields were normal; however, those records were not available. The anterior segments were normal. Ophthalmoscopy showed chorioretinal atrophy similar to that in his brother, but it had not progressed as far centrally.

Patient III/3. In this 70-year-old male patient, we performed a fundoscopic examination and serum examinations. The other clinical data are based on the notes of different ophthalmologists who examined this patient over the past 20 years. His general medical history was unremarkable, and he had no systemic disease that might have affected eye function. He did not receive any medication regularly.

Difficulties with visual function were noted when he was approximately 40 years old. At the age of 50 years, his visual acuity was 20/20 in both eyes. Visual fields showed concentric narrowing to approximately 15 degrees for the III/4 target of the Goldmann perimeter. He underwent cataract surgery with lens implantation in both eyes at age 63. Nd-YAG laser capsulotomy was performed 3 years later on the left eye. At the age of 64 years, his visual acuity was 20/40 in the right eye and 20/200 in the left. Six years later, his visual acuity was 20/60 in the right eye and 20/100 in the left. Visual fields tested with a Goldmann perimeter showed a remaining central island of 3 degrees for the III/4 target (Figure 2).

Fundus findings obtained by previous examiners showed retinal degeneration, but no detailed description was given. At the age of 70 years, the bare sclera was visible with a general loss of retinal pigment epithelium and choroidal vessels except for the foveal area. The optic disc was pale, and the retinal vessels were attenuated. There was no pigment clumping (Figure 3).

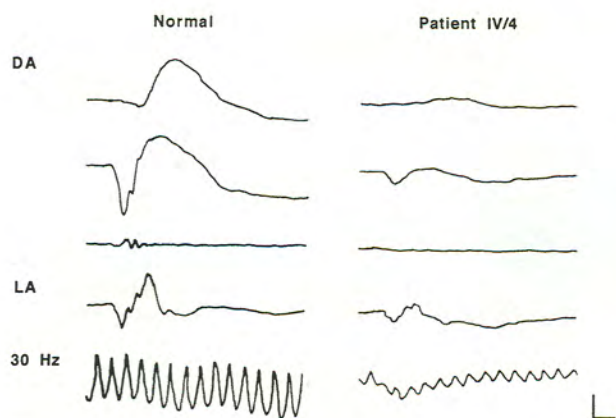


Fig. 4. Electrophysiological recordings of patient IV/4 (right) and a normal control subject (left). The upper three traces show recordings at dark adaptation (DA, calibration: $200 \mu\text{V}$ and 20 ms): rod response, mixed rod-cone response, and oscillatory potentials. The lower two traces show recordings at light adaptation (LA), cone response (calibration: $100 \mu\text{V}$ and 20 ms), and 30-Hz flicker (calibration: $100 \mu\text{V}$ and 50 ms).

degrees from the fovea showed elevated cone thresholds (0.8 and 1.3 log units above the mean for the right eye and the left eye, respectively) and rod thresholds (1.0 and 1.5 log units above the mean for the right eye and the left eye, respectively) bilaterally. Electroretinogram a- and b-waves were severely subnormal for cone- and rod-mediated responses. Cone and rod implicit times were prolonged (Figure 7). Electrooculography light-to-dark ratios were subnormal in each eye (right eye, 1.26 ; left eye, 1.16 ; normal, > 1.85).

The fasting serum ornithine level was normal ($116 \mu\text{mol/L}$; normal, $30\text{--}120 \mu\text{mol/L}$). The glutamine ($947 \mu\text{mol/L}$; normal, $410\text{--}690 \mu\text{mol/L}$) and alanine ($764 \mu\text{mol/L}$; normal, $210\text{--}660 \mu\text{mol/L}$) levels were elevated in one sample but normal on repeat testing. The arginine level ($135 \mu\text{mol/L}$) was at the upper limit of the normal range ($20\text{--}130 \mu\text{mol/L}$). All other amino acid levels were normal. Ornithine-delta-aminotransferase activity in

cultured skin fibroblasts was also normal, and the apparent Km for ornithine and α -ketoglutarate was within the normal range.

Case 2. This 70-year-old man reported slowly progressive, life-long limited side vision, but did not produce noticeable disability until the age of 65 years. Since the age of 30 years he noted progressively worse night vision. At age 63 his visual acuity measured $20/60$ in the right eye and $20/40$ in the left, attributed mostly to the presence of moderate cataracts in each eye. Several ophthalmologists believed that he might have an atypical form of retinitis pigmentosa. A nonGanzfeld ERG obtained when the patient was 64 years old demonstrated markedly subnormal photopic and scotopic responses. Before age 67 the patient drove a car, but he drove at night only when he had no other option. His only medication was aspirin for osteoarthritis, which he had taken since his third decade of life.

At 70 years of age his best corrected visual acuity, wearing $+2.25\text{--}2.50 \times 87^\circ$ in the right eye and $+2.25\text{--}1.25 \times 97^\circ$ in the left, was $20/200$ in the right eye and $20/100$ in the left. Moderately advanced nuclear sclerotic cataracts were present bilaterally; from the results of laser interferometry, visual acuity potentials of $20/40$ in the right eye and $20/75$ in the left were predicted. Fundus examination showed extensive peripheral chorioretinal atrophy bilaterally. Goldmann perimetry displayed enlarged blind spots, severe constriction of peripheral visual fields, and depression of the central retinal sensitivity bilaterally. Major crossing errors with no discernible axis were made with the Panel D-15; the patient refused to complete the 100-Hue test, claiming no visible difference in the color of the chips upon which to perform the match. Results of dark adaptometry tested with a Goldmann-Weekers adaptometer with a target size of 11 degrees in the diameter in the upper field 10 degrees from the fovea disclosed marked elevation of the cone threshold (2.0 log units above the mean in both eyes), no discernible cone-rod break, and marked elevation of the final rod dark adaptation threshold (2.8 and 3.5 log units above the mean in the right eye and left eye, respectively). Ganzfeld ERGs (Figure 7) revealed a profound loss of dark-adapted rod, dark-adapted cone, and light-adapted cone mediated amplitudes for a- and b-waves. Rod- and cone-mediated b-wave implicit times were prolonged. Essentially no light-induced rise of the resting potential was seen on the EOG (light-to-dark ratios of 1.00 in both eyes).

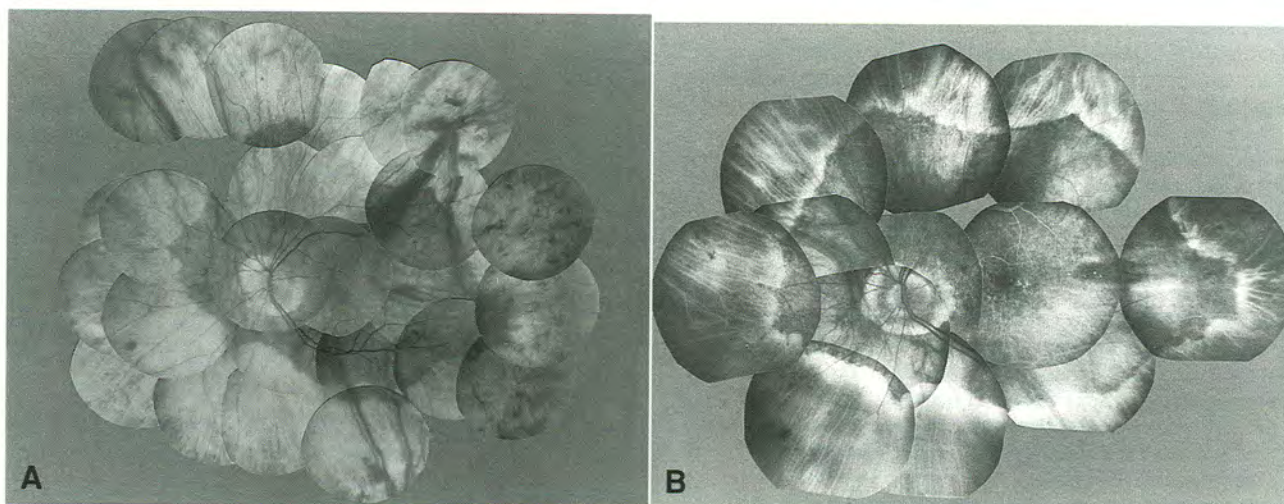


Fig. 5. Fundusoscopic appearance of simplex case 1 (81 years of age). **A**, Montage of fundus images of the left eye. **B**, Montage of the corresponding fluorescein angiogram.

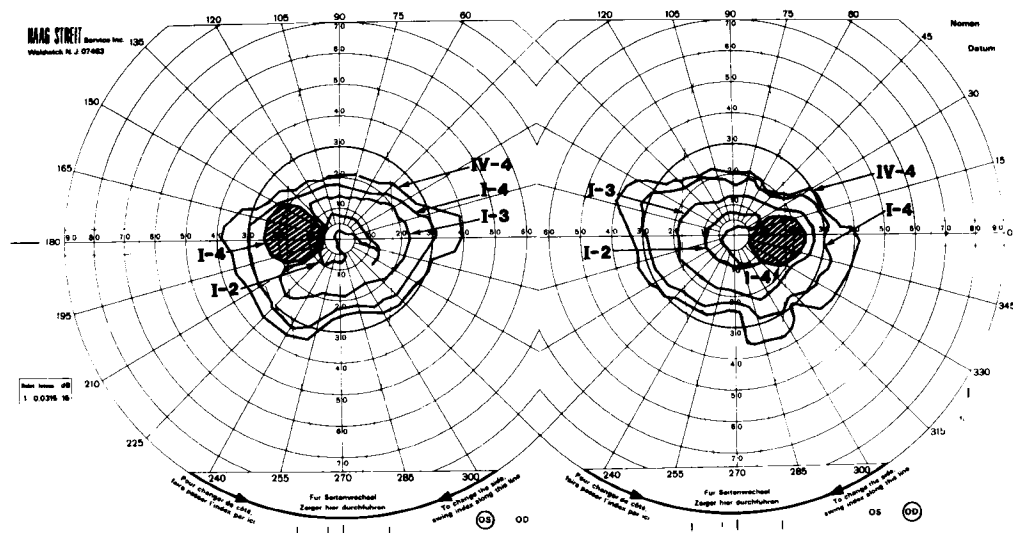


Fig. 6. Visual field defects in simplex case 1 (81 years of age).

At the age of 72 years, an intracapsular cataract extraction was performed on the right eye; a sample of aqueous humor was removed at the time of surgery and analyzed to determine amino acid levels. Postoperative visual acuity with aphakic correction was 20/70.

The levels of fasting serum ornithine (130 $\mu\text{mol/L}$) and arginine (139 $\mu\text{mol/L}$) were at the upper limit of the normal ranges. The level of ornithine in the aqueous humor was 42 $\mu\text{mol/L}$ (normal mean, 63; range, 38–84).²⁵ Ornithine-delta-aminotransferase activity in cultured skin fibroblasts was also normal, and the apparent K_m for ornithine and α -ketoglutarate was within the normal range.

Case 3. A 62-year-old man, first examined at the University of Illinois in December 1980 (G.A.F.), reported mild photosensitivity and decreased distance and near vision that had lasted 2 years. He denied having problems with color or night vision. His medications included methyl dopa 250 mg bid and hydrochlorothiazide 50 mg daily; both were to control systemic hypertension. One year earlier, he had undergone successful quadruple vessel coronary artery bypass surgery. Seven years earlier, an aortic aneurysm had been repaired with a dacron graft. There was no history of diabetes, neurologic disease, peptic ulcer disease, or renal disease. Family history showed no members with night blindness or any blinding disease. No other family members were available for examination.

Manifest refraction with +1.00 sphere in both eyes improved vision to 20/30 -2 in the right eye and to 20/30 +2 in the left. With a +1.50 add for near, the patient was able to read four-point size print. Although both anterior segments were unremarkable, trace cells in the right vitreous and +1 cells in the left vitreous were detected on slit-lamp examination. Dilated fundus examination showed typical findings of gyrate atrophy. There were extensive, atrophic, scalloped lesions circumferential within the midperipheral retina of each eye. The optic disc and retinal vessels appeared normal (Figure 8).

Fluorescein angiography showed characteristic atrophy of the retina and choroid in the sharply defined scalloped regions. Some of the large choroidal vessels were spared. The posterior pole of the retina showed very subtle, granular pigment epithelial window defects. Fluorescein stained the intact choriocapillaris at the border of the degenerative areas.

Goldmann visual fields showed marked peripheral constriction (Figure 9). Color vision with the Farnsworth-Munsell 100 Hue test showed a mild blue-yellow defect. Dark adapted thresholds were obtained with a Goldmann-Weekers dark adaptometer using a 2-degree test stimulus. Rod thresholds tested at 15 degrees superior and inferior to the fovea were at the upper limits of the normal range. At 30 degrees superior and inferior to the fovea, rod thresholds were elevated by 1 log unit, whereas at 45 degrees thresholds were elevated by 2 log units above the upper limit of the normal range.

Electroretinography showed undetectable cone responses to a single-flash stimulus and 30-Hz flicker cone responses that were reduced to 85% below the normal value. The dark-adapted, rod-dominant response showed a b-wave amplitude that was only 10% of the normal response. Isolated rod function to a single-flash, blue stimulus was undetectable.

Independent analysis of the patient's serum and urine by three separate laboratories showed normal ornithine levels (120 $\mu\text{mol/L}$) and normal levels for other amino acids. Ornithine ketoacid aminotransferase activity was normal.

Discussion

The differential diagnosis of inherited, progressive, generalized, choroidal dystrophies includes three entities: diffuse choriocapillaris atrophy, choroideremia, and gyrate atrophy.^{33–36} Diffuse choriocapillaris atrophy is a rare disorder with autosomal dominant inheritance.³³ Choriocapillaris atrophy and pigment clumping originate at the posterior pole and result in early visual loss. The disease slowly progresses to an end stage with large choroidal vessels visible as thin, white cords.

Choroideremia is an X-linked, recessive, choroidal dystrophy that begins in the periphery and progresses to the posterior pole.^{32,51} It is characterized by a scalloped loss of retinal pigment epithelium and choriocapillaris. Night blindness occurs at an

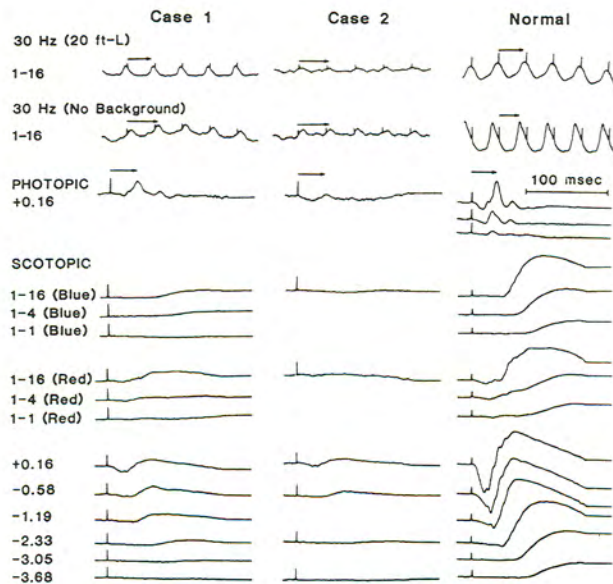


Fig. 7. Electoretinography (ERG) recordings of patients with simplex cases 1 and 2 compared with that of a normal control subject. The intensity settings for the Grass stimulator are to the left for the scotopic blue and red stimuli. The numbers to the left of the white-light waveforms indicate the integrated light intensity in log foot-lambert seconds. To act as an indicator of the vertical scale, the stimulus spikes for the 30-Hz flicker, photopic, and scotopic waveforms have been set at 10 μ V, 25 μ V, and 50 μ V for patients 1 and 2 and 75 μ V, 75 μ V, and 100 μ V for the normal ERG. Note that the 30-Hz flicker response without background is prolonged for both patients.

early stage, and severely constricted visual fields and visual loss are present in the patient's fifth decade of life. Choroideremia is caused by a deficiency of Rab geranylgeranyl transferase. The gene is located at Xq21.^{52,53}

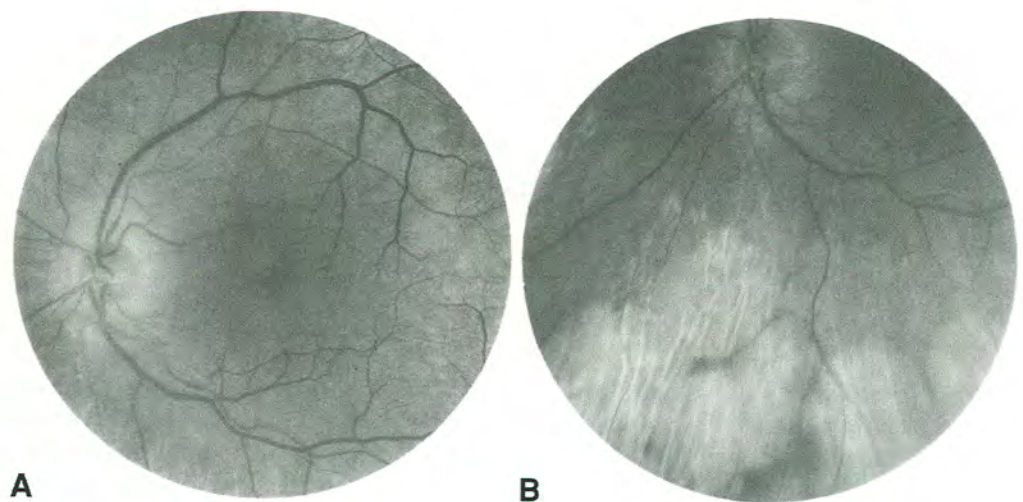
The patients in this study have a slowly pro-

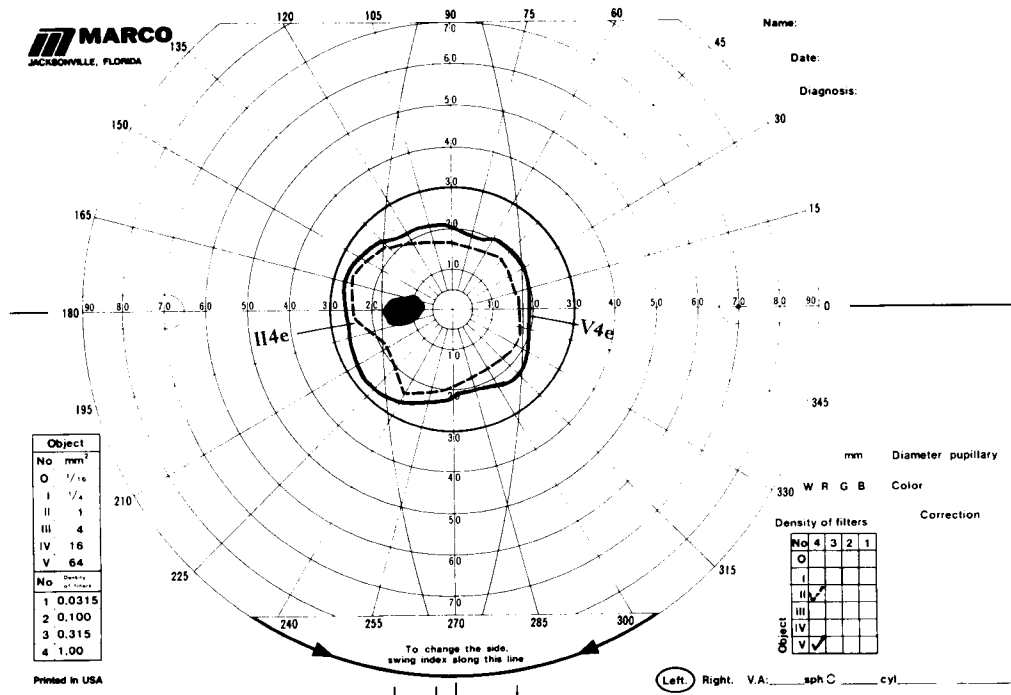
gressive choroidal dystrophy, which most resembles gyrate atrophy. With this disorder, the chorioretinal atrophy begins in the periphery and progresses more centrally with increasing age. This is in contrast to findings in patients with diffuse choriocapillaris atrophy, which originates at the posterior pole. The margins of the patches of chorioretinal atrophy are often sharply delineated in the early stages, and pigment clumping is rare. The most advanced form of the disease, which was present in patient III/3 (Family M), resembled a late stage of gyrate atrophy or choroideremia. None of our patients had hyperornithinemia, which excludes gyrate atrophy as a diagnosis. Moreover, the inheritance pattern in the family suggests an autosomal dominant trait, which would exclude autosomal recessive gyrate atrophy and X-linked choroideremia.

On ophthalmoscopy, the findings in these patients resembled gyrate atrophy, but in other ophthalmologic aspects there were differences. High myopia was not present in our patients. There was no early cataract formation, and cataract surgery was only performed in patients older than 63 years of age. The onset of subjective signs and the stage of legal blindness occurred later in life than they would in patients with gyrate atrophy. Functional loss as measured with ERG was milder in these patients than in patients with gyrate atrophy.

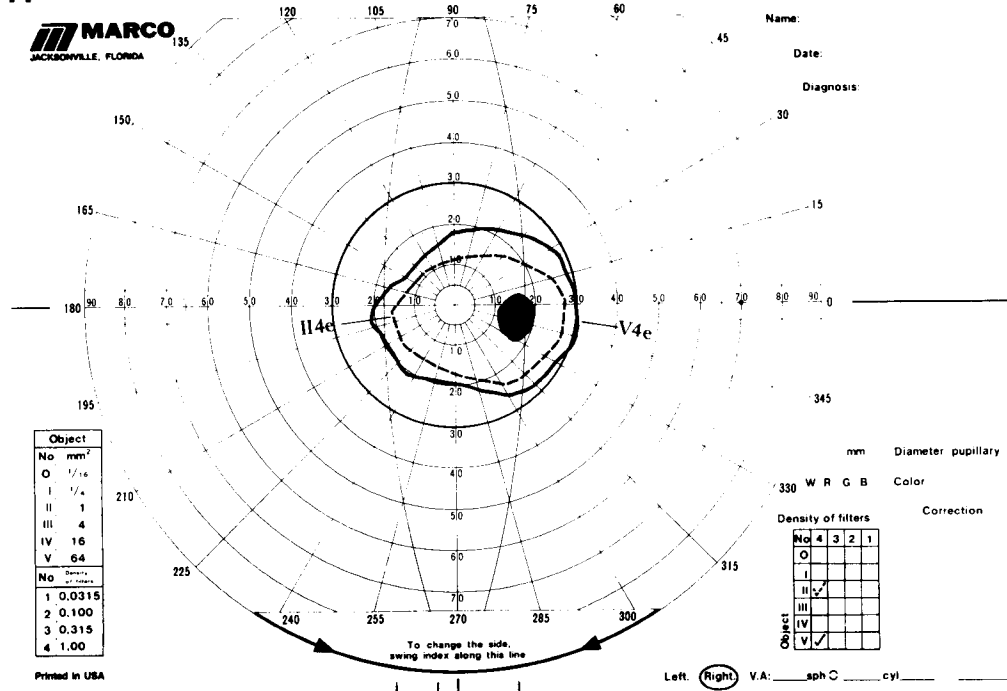
It is difficult to define whether the disease process was the same in the family and the simplex cases. In none of the simplex cases was the family history suggestive of other affected individuals, but information on other family members was

Fig. 8. Funduscopy appearance of simplex case 3 (62 years of age). **A**, Posterior pole of the left eye. **B**, Choroidal atrophy below the optic disc of the left eye.





A



B

Fig. 9. Visual field defects in simplex case 3 (62 years of age). **A,** Left eye. **B,** Right eye.

lacking. There was no consanguinity. The existence of a new mutation for an autosomal dominant disorder or the possibility that other family members were affected but not recognized cannot be excluded. There was no indication that the

peripheral changes were secondary to inflammation or uveitis. However, the functional loss was apparently more advanced in the older patient in family M than in the patients of similar age with the simplex cases of the disease. Only the 70-

year-old patient in family M had a nonrecordable ERG.

Biochemically, our patients are quite distinct from those with gyrate atrophy. Serum ornithine levels and OAT activity in cultured skin fibroblasts were normal. Further, the apparent K_m for ornithine and α -ketoglutarate was normal in the two patients in whom it was measured, eliminating the possibility that these patients had a variant OAT with deficient activity at low concentrations of substrates. It is important to distinguish the disorder in the patients described here from gyrate atrophy, because the latter is amenable to treatment in some cases, either by the administration of pyridoxine to vitamin B₆-responsive patients or by dietary restriction of arginine to non-B₆-responsive patients.^{6-11,54-61}

Other cases of gyrate atrophy phenotype without hyperornithinemia have been described. In four families, consanguineous marriages led to ophthalmoscopic findings similar to gyrate atrophy but associated with other defects, e.g., muscular dystrophy and mental retardation, iminoglycinuria and iris atrophy, nystagmus and strabismus, and congenital amaurosis.³⁷⁻⁴¹ In another family without consanguinity, choroidal dystrophy developed in two men, one of whom had spontaneous lens subluxation in both eyes.^{37,38} There are only a few cases that parallel the findings in the simplex cases observed in this study. Bargum⁴² observed a 67-year-old female patient with late-onset choroidal dystrophy and normal serum levels of ornithine. There was no family history of eye disease. The patient's six brothers and their children were examined and had normal ophthalmologic findings. Barcaroli et al⁴³ described three patients with choroidal dystrophy similar to gyrate atrophy who had normal serum ornithine levels and were emmetropic or hyperopic. In some cases of advanced retinitis pigmentosa, peripheral patches of choroidal atrophy may be seen, but the other ophthalmoscopic features of advanced retinitis pigmentosa enable a correct diagnosis to be made.¹³

Three families with members with autosomal dominant gyrate atrophy-like choroidal dystrophy have been described. Raitta et al⁶² observed a patient in whom spinocerebellar ataxia was an additional finding. Marchesani and Sautter⁴⁴ described a pedigree with several affected members in three generations; however, besides the pedigree and a fundus drawing, no further information was given. Krill and Archer³³ examined one female and two male siblings and their affected

mother. In their family, the area of choroidal dystrophy was variable, but the fovea was involved in a late stage of the disease. These two families are the only ones that showed dominant inheritance and that are similar to the family in the present report.

Although defects in the OAT gene are associated with gyrate atrophy, the precise pathogenesis of the choroidal dystrophy has not yet been defined, and different theories are under discussion.¹³ There are two possible explanations for the development of these choroidal dystrophies. First, there may be one single and yet undefined disease process that is common to our patients and patients with gyrate atrophy. A second explanation is that different disease processes may lead to a similar chorioretinal degeneration. The difference in severity of functional loss in patients with gyrate atrophy, family M, and the simplex cases suggests that different disease processes with a similar ophthalmoscopic picture are more likely. A recently described mouse model of gyrate atrophy may be helpful for the further analysis of the pathogenesis of these choroidal dystrophies.⁶³

Key words: choroid, electrooculogram, electroretinogram, gyrate atrophy, retinal degeneration.

References

1. Jacobsohn E. Ein Fall von Retinitis pigmentosa atypica. *Klin Monatsbl Augenheilk* 1888;26:202-206.
2. McCulloch C, Marliss EB. Gyrate atrophy of the choroid and retina with hyperornithinemia. *Am J Ophthalmol* 1975;80:1047-1057.
3. McCulloch C, Marliss EB. Gyrate atrophy of the choroid and retina: clinical ophthalmologic and biochemical considerations. *Trans Am Ophthalmol Soc* 1976;73:153-171.
4. Kaiser-Kupfer MI, Valle D, Del Valle LA. A specific enzyme defect in gyrate atrophy. *Am J Ophthalmol* 1978;85:200-204.
5. Kaiser-Kupfer MI, Valle D, Bron AJ. Clinical and biochemical heterogeneity in gyrate atrophy. *Am J Ophthalmol* 1980;89:219-222.
6. Kennaway NG, Weleber RG, Buist NR. Gyrate atrophy of the choroid and retina with hyperornithinemia: biochemical and histologic studies and response to vitamin B₆. *Am J Hum Genet* 1980;32:529-541.
7. Berson EL, Shih VE, Sullivan PL. Ocular findings in patients with gyrate atrophy on pyridoxine and low-protein low-arginine diets. *Ophthalmology* 1981;88:311-315.
8. Valle D, Walser M, Brusilow S, et al. Gyrate atrophy of the choroid and retina: biochemical considerations and experience with an arginine-restricted diet. *Ophthalmology* 1981;88:325-330.
9. Weleber RG, Kennaway NG, Buist NR. Gyrate atrophy of the choroid and retina: approaches to therapy. *Int Ophthalmol* 1981;4:23-32.
10. Hayasaka S, Saito T, Nakajima H, et al. Gyrate atrophy with

- hyperornithinaemia: different types of responsiveness to vitamin B₆. *Br J Ophthalmol* 1981;65:478–483.
11. Hayasaka S, Saito T, Nakajima H, et al. Clinical trials of vitamin B₆ and proline supplementation for gyrate atrophy of the choroid and retina. *Br J Ophthalmol* 1985;69:283–290.
 12. Kaiser-Kupfer MI, Ludwig IH, de Monasterio F, et al. Gyrate atrophy of the choroid and retina: early findings. *Ophthalmology* 1985;92:394–401.
 13. Weleber RG, Kennaway N. Gyrate atrophy of the choroid and retina. In: Heckenlively JR, ed. *Retinitis Pigmentosa*. Philadelphia: Lippincott, 1988;213–220.
 14. Inana G, Hotta Y, Zintz C, et al. Expression defect of ornithine aminotransferase gene in gyrate atrophy. *Invest Ophthalmol Vis Sci* 1988;29:1001–1005.
 15. Akaki Y, Hotta Y, Mashima Y, et al. A deletion in the ornithine aminotransferase gene in gyrate atrophy. *J Biol Chem* 1992;267:12950–12954.
 16. Mashima Y, Murakami A, Weleber RG, et al. Nonsense-codon mutations of the ornithine aminotransferase gene with decreased levels of mutant mRNA in gyrate atrophy. *Am J Hum Genet* 1992;51:81–91.
 17. Mashima Y, Weleber RG, Kennaway NG, Inana G. A single-base change at a splice acceptor site in the ornithine aminotransferase gene causes abnormal RNA splicing in gyrate atrophy. *Hum Genet* 1992;90:305–307.
 18. Brody LC, Mitchell GA, Obie C, et al. Ornithine-d-aminotransferase mutations causing gyrate atrophy: allelic heterogeneity and functional consequences. *J Biol Chem* 1992;267:3302–3307.
 19. Ramesh V, McClatchey AI, Ramesh N, et al. Molecular basis of ornithine aminotransferase deficiency in B₆-responsive and nonresponsive forms of gyrate atrophy. *Biochemistry* 1988;85:3777–3780.
 20. Mitchell GA, Brody LC, Sipila I, et al. At least two mutant alleles of ornithine d-aminotransferase cause gyrate atrophy of the choroid and retina in Finns. *Proc Natl Acad Sci USA* 1989;86:197–201.
 21. Michaud J, Thompson GN, Brody LC, et al. Pyridoxine-responsive gyrate atrophy of the choroid and retina: clinical and biochemical correlates of the mutation A226V. *Am J Hum Genet* 1995;56:616–622.
 22. Kobayashi T, Ogawa H, Kasahara M, Shiozawa Z, Matsuzawa T. A single amino acid substitution within the mature sequence of ornithine aminotransferase obstructs mitochondrial entry of the precursor. *Am J Hum Genet* 1995;57:284–291.
 23. Simell O, Takki K. Raised plasma-ornithine and gyrate atrophy of the choroid and retina. *Lancet* 1973;1:1031–1033.
 24. Takki K, Simell O. Genetic aspects in gyrate atrophy of the choroid and retina with hyperornithinaemia. *Br J Ophthalmol* 1974;58:907–916.
 25. Takki K. Gyrate atrophy of the choroid and retina associated with hyperornithinaemia. *Br J Ophthalmol* 1974;58:3–23.
 26. Kennaway NG, Weleber RG, Buist NR. Gyrate atrophy of the choroid and retina: deficient activity of ornithine ketoacid aminotransferase in cultured skin fibroblasts. *N Engl J Med* 1977;297:1180.
 27. O'Donnell JJ, Sandman RP, Martin SR. Deficient L-ornithine: 2-oxoacid aminotransferase activity in cultured fibroblasts from a patient with gyrate atrophy of the retina. *Biochem Biophys Res Commun* 1977;79:396–399.
 28. Trijbels JM, Sengers RC, Bakkeren JA, et al. L-ornithine-ketoacid-transaminase deficiency in cultured fibroblasts of a patient with hyperornithinaemia and gyrate atrophy of the choroid and retina. *Clin Chim Acta* 1977;79:371–377.
 29. Valle D, Kaiser KM, Del VL. Gyrate atrophy of the choroid and retina: deficiency of ornithine aminotransferase in transformed lymphocytes. *Proc Natl Acad Sci USA* 1977;74:5159–5161.
 30. O'Donnell JJ, Sandman RP, Martin SR. Gyrate atrophy of the retina: inborn error of L-ornithine:2-oxoacid aminotransferase. *Science* 1978;200:200–201.
 31. Shih VE, Berson EL, Mandell R, Schmidt SY. Ornithine ketoacid transaminase deficiency in gyrate atrophy of the choroid and retina. *Am J Hum Genet* 1978;30:174–179.
 32. Mauthner L. Ein Fall von Choroideremia. *Berichte der Naturwissenschaftlichen und Medizinischen Vereinigung Innsbruck* 1871;2:191.
 33. Krill AE, Archer D. Classification of the choroidal atrophies. *Am J Ophthalmol* 1971;72:562–585.
 34. Hayasaka S, Shoji K, Kanno C, et al. Differential diagnosis of diffuse choroidal atrophies: diffuse choriocapillaris atrophy choroideremia and gyrate atrophy of the choroid and retina. *Retina* 1985;5:30–37.
 35. Jiménez-Sierra JM, Ogden TE, van Boemel GB. *Inherited Retinal Diseases: A Diagnostic Guide*. St. Louis: Mosby, 1989;17–33.
 36. Weleber RG. Degeneration and atrophy of the choroid. In: Tasman W, Jaeger EA, eds. *Duane's Clinical Ophthalmology*, vol. 4, ch. 66. Philadelphia: Lippincott-Raven, 1995.
 37. Jaeger W, Kettler JV, Hildorf C, Lutz P. Gibt es verschiedene Typen der Atrophia gyrata chorioideae et retinae (Atrophia gyrata chorioideae et retinae mit und ohne Ornithinämie). *Ber Zusammenkunft Dtsch Ophthalmol Ges* 1978;75:655–663.
 38. Jaeger W, Kettler JV, Lutz P, Hildorf C. Differential diagnosis of gyrate atrophy of the choroid and retina. *Metabolic Pediatric Ophthalmology* 1978;3:189–191.
 39. Hayasaka S, Mizuno K, Yabata K, et al. Atypical gyrate atrophy of the choroid and retina associated with iminoglycinuria. *Arch Ophthalmol* 1982;100:423–425.
 40. Mizuno K, Hayasaka S, Tsuchiya M. Leber's congenital amaurosis in a patient with normal amino acid metabolism and fundus features characteristic of gyrate atrophy. *Am J Ophthalmol* 1983;96:806–807.
 41. Mishima H, Hirata H, Ono H, et al. A Fukuyama type of congenital muscular dystrophy associated with atypical gyrate atrophy of the choroid and retina: a case report. *Acta Ophthalmol (Copenh)* 1985;63:155–159.
 42. Bargum R. Differential diagnosis of normoornithinaemic gyrate atrophy of the choroid and retina. *Acta Ophthalmol (Copenh)* 1986;64:369–373.
 43. Barcaroli M, Forte R, Iacobelli L, et al. Gyrate atrophy: atypical findings and differential diagnosis. *Invest Ophthalmol Vis Sci* 1996;37(Suppl):S505.
 44. Marchesani O, Sautter H. *Atlas des Augenhintergrundes*. München: Urban & Schwarzenberg, 1957:197–198.
 45. Marmor MF, Arden GB, Nilsson SEG, Zrenner E. Standard for clinical electroretinography. *Arch Ophthalmol* 1989;107:816–819.
 46. Marmor MF, Zrenner E. Standard for clinical electrooculography. *Arch Ophthalmol* 1993;111:601–604.
 47. Fishman GA, Buckmann G, van Every T. *Fundus flavimaculatus: a clinical classification*. *Doc Ophthalmol Proc Ser* 1977;13:213–220.
 48. Fishman GA, Fishman M, Maggiano J. Macular lesions as-

- sociated with retinitis pigmentosa. *Arch Ophthalmol* 1977;95:798–803.
49. Kellner U, Bornfeld N, Foerster MH. Severe course of cutaneous melanoma associated retinopathy. *Br J Ophthalmol* 1995;79:746–752.
 50. Weleber RG, Eisner A. Retinal function and physiological studies. In: Newsome DA, ed. *Retinal Degeneration*. New York: Raven Press, 1987;21–69.
 51. Heckenlively JR, Bird AC. Choroideremia. In: Heckenlively JR, ed. *Retinitis Pigmentosa*. Philadelphia: Lippincott, 1988;176–187.
 52. Cremers FPM, van de Pol DJR, van Kerkhoff LPM, et al. Cloning of a gene that rearranges in patients with choroideremia. *Nature* 1990;347:674–677.
 53. Seabra MC, Brown MS, Goldstein JL. Retinal degeneration in choroideremia: deficiency of Rab geranylgeranyl transferase. *Science* 1993;259:377–381.
 54. McInnes RR, Arshinoff SA, Bell L, McCulloch C. Treatment of gyrate atrophy of the choroid and retina with a low arginine diet. *Trans Am Ophthalmol Soc* 1980;78:226–242.
 55. McInnes RR, Arshinoff SA, Bell L, et al. Hyperornithinaemia and gyrate atrophy of the retina: improvement of vision during treatment with a low-arginine diet. *Lancet* 1981;1:513–516.
 56. Kaiser-Kupfer MI, de Monasterio F, Valle D, et al. Visual results of a long-term trial of a low-arginine diet in gyrate atrophy of choroid and retina. *Ophthalmology* 1981;88:307–310.
 57. Shih VE, Berson EL, Gargiulo M. Reduction of hyperornithinemia with a low protein low arginine diet and pyridoxine in patients with a deficiency of ornithine-ketoacid transaminase (OKT) activity and gyrate atrophy of the choroid and retina. *Clin Chim Acta* 1981;113:243–251.
 58. Tada K, Saito T, Omura K, et al. Hyperornithinaemia associated with gyrate atrophy of the choroid and retina: *in vivo* and *in vitro* response to vitamin B₆. *J Inher Metab Dis* 1981;4:61–62.
 59. Weleber RG, Kennaway NG. Clinical trial of vitamin B₆ for gyrate atrophy of the choroid and retina. *Ophthalmology* 1981;88:316–324.
 60. Berson EL, Hanson AD, Rosner B, Shih VE. A two-year trial of low protein low arginine diets or vitamin B₆ for patients with gyrate atrophy. *Birth Defects* 1982;18:209–218.
 61. Kaiser-Kupfer MI, Caruso RC, Valle D. Gyrate atrophy of the choroid and retina: long-term reduction of ornithine slows retinal degeneration. *Arch Ophthalmol* 1991;109:1539–1548.
 62. Raitta C, Koskinen L, Simell O. Progressive retino-choroidal atrophy with spinocerebellar ataxia. *Neuro-Ophthalmol* 1984;4:1–19.
 63. Wang T, Lawler AM, Steel G, et al. Mice lacking ornithine-delta-aminotransferase have paradoxical neonatal hypoor-nithinaemia and retinal degeneration. *Nat Genet* 1995;11:185–190.